

to III and is assigned the structure IX. Copper-catalyzed decomposition of V in cyclohexene at 130° gave VII (43%); no evidence for the formation of a norcarane derivative could be obtained. Copper-catalyzed decomposition of I in either ethanol or cyclohexene gave II (91 and 94%, respectively).

We consider that these results are best interpreted in the following manner. In the decomposition of diazoisofenchone, a carbene (or carbene-copper complex⁹) is formed which in benzene or cyclohexene rearranges to VII by concerted migration of the *endo* C₅ methyl group and C₃-C₅ bond formation; in ethanol, reaction of the carbene with the solvent takes precedence over its rearrangement, giving IX. In the decomposition of diazocamphor, loss of nitrogen may be concerted with 1,3-hydrogen migration and C₃-C₅ bond formation, giving II directly.¹⁰

On this view, the migration of the methyl group in the formation of VII may be classed as intermediate between a 1,2 and 1,3 alkyl shift. The formation of VII from the carbene may equivalently be viewed as an insertion reaction of the carbene at the *endo* C₅-CH₃ bond. No simple insertion reactions of carbenes at carbon-carbon single bonds have been described,¹¹ although certain other rearrangement reactions of carbenes may be viewed as proceeding in this fashion.¹²

(9) P. Yates, *J. Am. Chem. Soc.*, **74**, 5376 (1952).

(10) W. Reusch, M. W. DiCarlo and L. Traynor, *J. Org. Chem.*, **26**, 1711 (1961), have shown that the related transformation of camphor hydrazone to tricyclene proceeds *via* intramolecular hydrogen transfer.

(11) Cf., however, W. von E. Doering and M. Jones, unpublished work referred to by W. Kirmse, *Angew. Chem.*, **73**, 161 (1961).

(12) Cf. L. Friedman and H. Schechter, *J. Am. Chem. Soc.*, **83**, 3159 (1961); M. S. Newman and A. Arkell, *J. Org. Chem.*, **24**, 385 (1959).

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CHEMISTRY OF MYCINOSE: 6-DEOXY-2,3-DI-O-METHYL-D-ALLOSE

Sir:

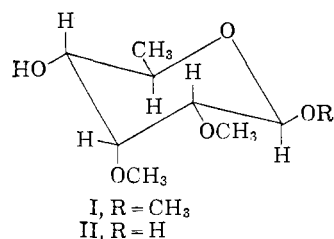
Methanolysis of chalconmycin¹ yields methyl chalcoside² and methyl mycinoside I, m.p. 88–88.5°, $[\alpha]_D^{25} -36^\circ$ (*c* 1.6%, chloroform) [*Anal.* Calcd. for C₉H₁₈O₅: C, 52.41; H, 8.80; O, 38.79; OCH₃(3), 45.1; C-CH₃(1), 7.3. Found: C, 52.38; H, 8.80; O, 39.07; OCH₃, 45.2; C-CH₃, 8.13].

Aqueous hydrolysis of I gives crystalline mycinoside II, m.p. 102–106° $[\alpha]_D^{25} -46^\circ \rightarrow -42^\circ$ (2 min.) $\rightarrow -36^\circ$ (30 min.) $\rightarrow -29^\circ$ (4 hr. and 24 hr.) (*c* 1.56%, water) [*Anal.* Calcd. for C₈H₁₆O₅: C, 49.99; H, 8.39; OCH₃(2), 32.29; C-CH₃(1), 7.82. Found: C, 49.94; H, 8.73; OCH₃, 31.98; C-CH₃, 7.42].

Mycinoside is oxidized with bromine water to give a crystalline lactone, m.p. 134–135°, which

(1) Parke, Davis & Company, Belgian Patent 587,213, August 2, 1960.

(2) P. W. K. Woo, H. W. Dion and Q. R. Bartz, *J. Am. Chem. Soc.*, **83**, 3352 (1961).



exhibits no maximum in the ultraviolet and shows sharp bands at 2.90 and 5.68 μ in the infrared spectrum. [*Anal.* Calcd. for C₈H₁₄O₅: C, 50.52; H, 7.42; OCH₃(2), 32.63; C-CH₃(1), 7.90; mol. wt., 190.19. Found: C, 50.36; H, 7.69; OCH₃, 32.24; C-CH₃, 7.72; mol. wt. (by titration), 182.] Mycinoside gives a positive iodoform test, reduces Fehling solution, and yields a pinkish-brown color with aniline hydrogen phthalate on papergrams; it does not reduce periodate, but the sodium borohydride reduction product takes up one mole to give acetaldehyde (0.75 mole as the 2,4-DNP) and a tetrose which was subsequently oxidized with nitric acid to give *meso*-dimethoxysuccinic acid, m.p. 162–163° [*Anal.* Calcd. for C₆H₁₀O₆: C, 40.45; H, 5.66; neut. equiv., 89.07. Found: C, 40.25; H, 5.83; neut. equiv., 91] which was identical with a synthetic sample (mixed m.p., *R_f* values, infrared spectrum). These data establish the structure of mycinoside as a 6-deoxy-2,3-di-O-methylhexose.

Treatment of I with boron trichloride³ for ten days at 4° and subsequent aqueous acid hydrolysis of the reaction product yields a crystalline sugar, m.p. 146–148°, $[\alpha]_D^{25} -4.7^\circ$ (7 min.) $\rightarrow 0^\circ$ (40 min. and 3 hr.) (*c* 3%, water) [*Anal.* Calcd. for C₆H₁₂O₅: C, 43.90; H, 7.37. Found: C, 44.18; H, 7.60], which was compared with a sample of 6-deoxy- β -D-allose prepared from digitoxin⁴ and found to be identical (mixed m.p., paper chromatography and paper electrophoresis,⁵ infrared spectrum, X-ray powder diagram). The phenylosazones⁴ (m.p. 180–183°, $[\alpha]_D^{25} -72^\circ$ [*c* 0.6%, pyridine-ethanol 2:3]) of the above two sugar samples were also identical.

Exhaustive methylation of I with methyl iodide and silver oxide, then nitric acid oxidation, esterification with methanol in the presence of hydrogen chloride, and finally treatment with methylamine, gives *ribo*-2,3,4-trimethoxy-N,N'-dimethylglutaramide,⁶ m.p. 147–147.5°, $[\alpha]_D^{25} 0^\circ$ (*c* 2%, chloroform) [*Anal.* Calcd. for C₁₀H₂₀O₅N₂: C, 48.37; H, 8.12; N, 11.28. Found: C, 48.47; H, 8.18; N, 11.50]. Thus, from the above, methyl mycinoside must be in the pyranose form as in I. Partial methylation of mycinoside with methyl iodide and silver oxide gives I. Thus, mycinoside probably exists in the pyranose form⁷ as in II.

(3) S. Allen, T. G. Bonner, E. J. Bourne and N. M. Saville, *Chem. & Ind.*, 630 (1958).

(4) F. Micheel, *Ber.*, **63**, 347 (1930).

(5) A. P. MacLennan and H. M. Randall, *Anal. Chem.*, **31**, 2020 (1959).

(6) P. A. Levene and J. Compton, *J. Biol. Chem.*, **116**, 183 (1936).

(7) N.m.r. spectroscopy shows that mycinoside does not mutarotate in chloroform until acid is added. It is therefore reasonable to assume that, under the methylation conditions in methyl iodide, it also does not mutarotate.

The n.m.r. spectra of I and II exhibit axial-axial splitting for H₁ and H₂ (I, $J_{1a,2a} = 7.6$ cps.; II, $J_{1a,2a} = 8.6$ cps.). The resulting absolute configuration at C-1, as indicated in II, agrees with that predicted, according to Hudson's rules, for a sugar in the D-series which mutarotates to a more positive value (*i.e.*, a β -D sugar).⁸

Thus, the second sugar obtained as a degradation product of chalcomycin has been isolated as 6-deoxy-2,3-di-O-methyl- β -D-allose.

(8) C. S. Hudson, "Advances in Carbohydrate Chem.," **3**, 15 (1948).

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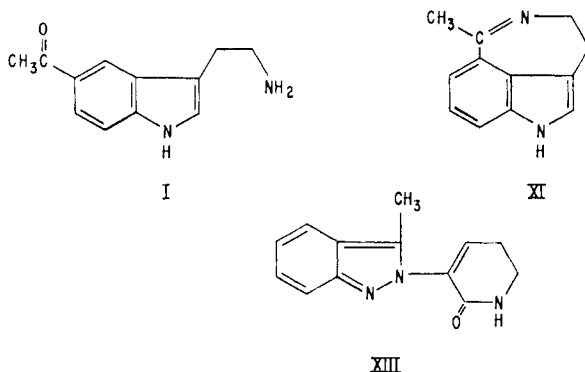
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RECEIVED DECEMBER 22, 1961

ACYLTRYPTAMINES. I. 5-ACETYLTRYPTAMINE AND RELATED COMPOUNDS

Sir:

The interest in 5-hydroxytryptamine (serotonin) and the total synthesis of reserpine has resulted in the preparation of a multiplicity of tryptamines for use in biological and synthetic investigations. The results of our efforts in another field led to our preparation of a series of novel tryptamines having acyl groups in the benzene ring. The discovery of the unusual hypotensive¹ properties of 5-acetyltryptamine (I) and some interesting chemical transformations during this extensive synthetic program has prompted us to publish some of our findings at this time.



The Japp-Klingemann reaction of *p*-acetylbenzenediazonium chloride with 2-oxopiperidine-3-carboxylic acid² gave the corresponding 3-(*p*-acetylphenyl)hydrazone of 2,3-piperidinedione (II, m.p. 229–231°, $\lambda_{\text{max}}^{\text{EtOH}}$ 238 m μ , ϵ 10,800, 351 m μ , ϵ 42,000. Calcd. for C₁₃H₁₅N₃O₂: C, 63.66; H, 6.16; N, 17.13. Found: C, 63.40; H, 6.28; N, 16.93). Refluxing II with 88% formic acid for four hours gave 6-acetyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-1-one (III; m.p. 370–372°, $\lambda_{\text{max}}^{\text{EtOH}}$ 273 m μ , ϵ 42,250, 302 m μ , ϵ 8,800. Calcd. for C₁₃H₁₂N₂O₂: C, 68.40; H, 5.30; N, 12.27. Found: C, 68.35; H, 5.33; N, 12.08). Hydrolysis³ of III in refluxing

(1) We thank Dr. Max Ben of the Department of Pharmacology for the observation of this hypotensive activity in the dog.

(2) R. A. Abramovitch and D. Shapiro, *Chem. and Ind.*, 1255 (1955); *J. Chem. Soc.*, 4589 (1956).

2 *N* KOH in 60% aqueous ethanol gave 5-acetyl-2-carboxytryptamine (IV, m.p. 337–343°, $\lambda_{\text{max}}^{\text{EtOH}}$ 266.5 m μ , ϵ 47,000, 303 m μ , ϵ 8,000. Calcd. for C₁₃H₁₄N₂O₃·H₂O: C, 59.08; H, 6.10; N, 10.60. Found: C, 59.04; H, 6.18; N, 10.77). Decarboxylation of IV in refluxing 20% hydrochloric acid containing 30% by volume glacial acetic acid gave 5-acetyltryptamine (I, m.p. 140–142°, $\lambda_{\text{max}}^{\text{EtOH}}$ 254 m μ , ϵ 34,400, 299 m μ , ϵ 7,850. Calcd. for C₁₂H₁₄N₂O: C, 71.26; H, 6.98; N, 13.85. Found: C, 70.99; H, 6.93; N, 14.09).

Reaction of *m*-acetylbenzenediazonium chloride with 2-oxopiperidine-3-carboxylic acid gave the 3-(*m*-acetylphenyl)hydrazone of 2,3-piperidinedione (V, m.p. 204–206°, $\lambda_{\text{max}}^{\text{EtOH}}$ 233 m μ , ϵ 19,500, 312.5 m μ , ϵ 21,200. Calcd. for C₁₃H₁₅N₃O₂·0.5H₂O: C, 61.40; H, 6.34; N, 16.53. Found: C, 61.58; H, 6.59; N, 16.87). Refluxing V with 88% formic acid gave a mixture of 5-acetyl and 7-acetyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-1-ones which were separated by fractional crystallization from absolute ethanol. The less soluble 7-acetyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-1-one (VI) had the m.p. 283–286°, $\lambda_{\text{max}}^{\text{EtOH}}$ 248 m μ , ϵ 27,100, 315 m μ , ϵ 23,700. Calcd. for C₁₃H₁₂N₂O₂: C, 68.40; H, 5.30; N, 12.27. Found: C, 68.30; H, 5.38; N, 12.45. The more soluble 5-acetyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-1-one (VII) had the m.p. 241–243°, $\lambda_{\text{max}}^{\text{EtOH}}$ 223 m μ , ϵ 24,500, 258 m μ , ϵ 11,800, 327 m μ , ϵ 10,090. Calcd. for C₁₃H₁₂N₂O₂: C, 68.40; H, 5.30; N, 12.27. Found: C, 68.62; H, 5.48; N, 12.06.

Hydrolysis of VI in refluxing 2 *N* KOH in 60% aqueous ethanol gave 6-acetyl-2-carboxytryptamine (VIII, m.p. 240–243°, $\lambda_{\text{max}}^{\text{EtOH}}$ 249 m μ , ϵ 25,300, 309 m μ , ϵ 21,700. Calcd. for C₁₃H₁₄N₂O₃·H₂O·0.5-C₂H₅OH: C, 58.52; H, 6.66; N, 9.75. Found: C, 58.74; H, 6.40; N, 9.73). Decarboxylation of VIII in refluxing 20% hydrochloric acid gave 6-acetyltryptamine⁴ (IX, m.p. 148–150°, $\lambda_{\text{max}}^{\text{EtOH}}$ 233 m μ (shoulder), ϵ 15,500, 253 m μ , ϵ 21,400, 301 m μ , ϵ 12,900. Calcd. for C₁₂H₁₄N₂O: C, 71.26; H, 6.98; N, 13.85. Found: C, 71.38; H, 7.18; N, 14.08).

Hydrolysis of VII in refluxing 2 *N* KOH in 60% aqueous ethanol gave 4-acetyl-2-carboxytryptamine (X, m.p. 314–317°, $\lambda_{\text{max}}^{\text{EtOH}}$ 216 m μ , ϵ 30,000, 245–52 m μ (shoulder), ϵ 13,900, 260 m μ , ϵ 15,000. Calcd. for C₁₃H₁₄N₂O₃: C, 63.40; H, 5.73; N, 11.38. Found: C, 63.59; H, 5.99; N, 11.46). Decarboxylation of X in refluxing 20% hydrochloric acid containing 30% by volume glacial acetic acid gave 3,4-dihydro-6-methyl-1H-azepino[5,4,3-cd]indole⁵ (XI, m.p. 272–277°, $\lambda_{\text{max}}^{\text{EtOH}}$ 241 m μ , ϵ 18,250. Calcd. for C₁₂H₁₂N₂: C, 78.22; H, 6.57; N, 15.21. Found: C, 77.98; H, 6.59; N, 15.36).

Reaction of *o*-acetylphenyldiazonium chloride with 2-oxo-piperidine-3-carboxylic acid gave the 3-(*o*-acetylphenyl)hydrazone of 2,3-piperidinedione (XII, m.p. 231–234°, $\lambda_{\text{max}}^{\text{EtOH}}$ 230 m μ , ϵ 17,300, 254 m μ , ϵ 11,700, 312.5 m μ , ϵ 14,700, 369 m μ , ϵ 13,200.

(3) S. Keimatsu, S. Sugawara and G. Kasuya, *J. Pharm. Soc. (Japan)*, **48**, 762 (1928); *Chem. Abstr.*, **23**, 834 (1929).

(4) This compound showed pressor activity in the dog, according to Dr. M. Osborne of the Department of Pharmacology.

(5) This compound represents the first of a series of derivatives of the novel ring system 1H-azepino[5,4,3-cd]indole.